



The role of endothelins in regulating bovine granulosa cell function

J. E. Ervin, L. F. Schultz and L. J. Spicer

^aDepartment of Animal Science, Oklahoma State University, Stillwater, OK 74078



INTRODUCTION

- Ovarian steroidogenesis and steroid-mediated signaling are critical for normal ovarian processes such as follicle growth and ovulation (1).
- Endothelin 1 (ET-1) and endothelin 2 (ET-2) are isoforms of vasoactive 21-amino acid peptides known to affect steroidogenesis within the ovary in several species (2,3,4,5).
- The role of endothelins in bovine ovarian steroidogenesis is inconclusive, but based on studies in other species we hypothesize that ET-1 and ET-2 inhibit bovine granulosa cell steroid production.
- This study is designed to investigate the effects of endothelins on bovine granulosa cells.

MATERIAL & METHODS

- Cell Culture:** Bovine ovaries were obtained from a local slaughterhouse. Follicular fluid was aspirated from small (1-5mm) follicles, and granulosa cells were isolated and exposed to various treatments (ET-1, ET-2, or ET-1+ET-2 with or without FSH or IGF1) in multiple experiments. FSH and IGF1 were treated at 30 ng/mL.
- RIA and Cell Counting:** In replicated experiments, culture medium was removed and analyzed for steroid production via radioimmunoassays. Granulosa cells were harvested with trypsin and counted using a Coulter Particle Counter.
- qPCR:** Cellular RNA was extracted & quantified using real-time PCR (18 S rRNA as housekeeping gene).
- Statistical Analysis:** Statistical analyses were performed using ANOVA and the general linear models (GLM) procedure of SAS for Windows (version 9.3, SAS Institute Inc., Cary, NY). If a significant main effect was identified in the ANOVA, then mean differences were determined by Fisher's protected least significant differences test (6). The values were reported as the least squares means \pm SEM.

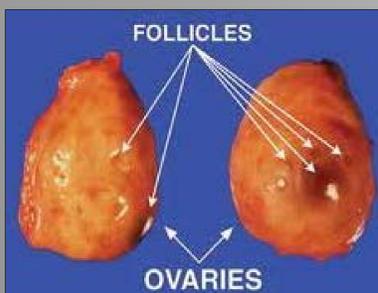


Figure 1: Bovine ovaries showing antral follicles.

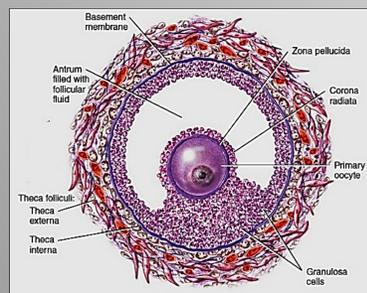


Figure 2: Cross-section of mature Graafian follicle showing oocyte, theca and granulosa cells.

RESULTS

Steroidogenesis

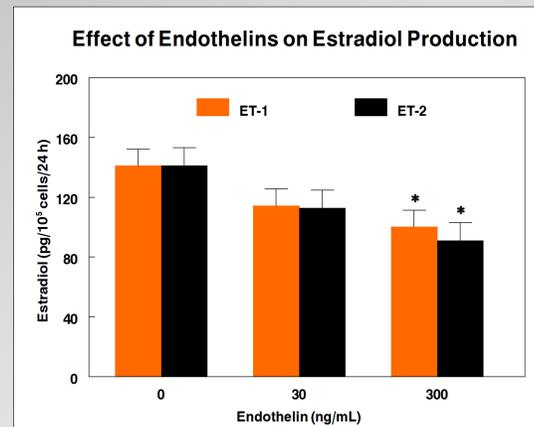


Figure 3: Dose response to endothelins in FSH plus IGF1-treated granulosa cells. * Means $P < 0.05$ compared to control.

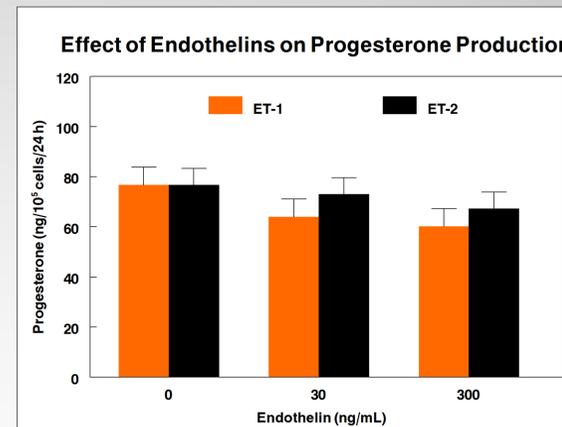


Figure 4: Dose response to endothelins in FSH plus IGF1-treated granulosa cells. Means did not differ $P > 0.05$.

Gene Expression

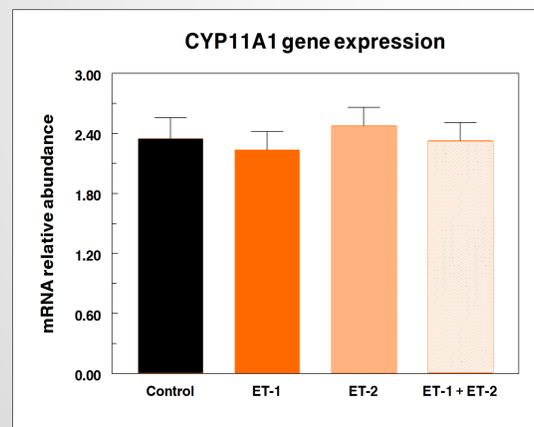


Figure 5: CYP11A1 response to various treatments in FSH plus IGF1-treated granulosa cells. Means did not differ $P > 0.05$.

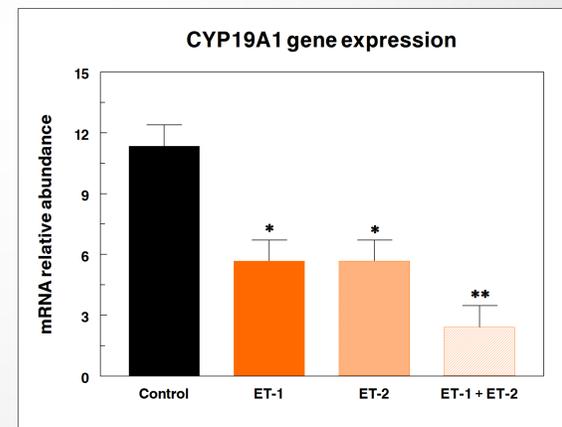


Figure 6: CYP19A1 response to various treatments in FSH plus IGF1-treated granulosa cells. *Means $P < 0.01$ compared to control. **Mean $P < 0.01$ compared to all other means.

SUMMARY & CONCLUSIONS

Steroidogenesis

- Both ET-1 and ET-2 inhibited estradiol production in a dose-dependent manner.
- Neither ET-1 nor ET-2 affected progesterone production.

Gene Expression

- CYP11A1 mRNA abundance was not affected by ET-1, ET-2 or ET-1+ET-2.
- CYP19A1 mRNA abundance was down-regulated by ET-1, ET-2 and ET-1+ET-2.

Conclusions

Endothelins suppress estradiol production by bovine granulosa cells, suggesting a role for endothelins in bovine ovarian function. Further investigation is necessary to fully understand the function of ET-1 and ET-2 in the bovine ovary and their role in regulating reproductive efficiency.

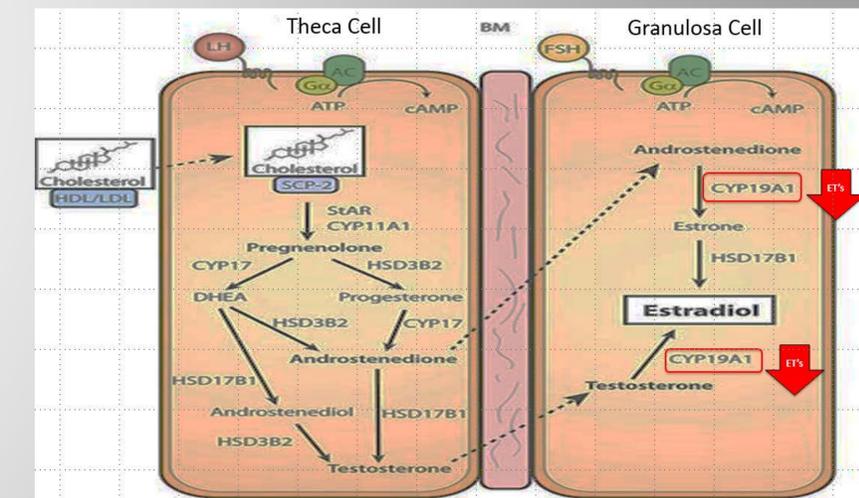


Figure 7: Enzymatic control of steroidogenesis in the growing follicle.

RESULTS

- Estradiol production was inhibited by 300 ng/mL of ET-1 ($P < 0.05$) and ET-2 ($P < 0.001$) (Fig. 3). While the 30 ng/mL treatments caused intermediate effects, they were not significantly different from controls (0 ng/mL) ($P > 0.05$).
- ET-1 and ET-2 did not affect progesterone production at either dosage ($P > 0.05$) (Fig. 4).
- Gene expression for side-chain cleavage enzyme (CYP11A1) was unaltered by ET-1, ET-2 or ET-1+ET-2 when compared to the control (Fig. 5). Gene expression for aromatase (CYP19A1) was inhibited by ET-1 ($P \leq 0.01$), ET-2 ($P \leq 0.01$) and ET-1+ET-2 ($P < 0.01$) (Fig. 6).

REFERENCES

- Jamnongjit, M., et al. Cell Cycle 5: 1178-1183, 2006.
- Yanagisawa, M., et al. Nature 332:411-415, 1998.
- Tedeschi, C., et al. Biol. Reprod. 51:1058-1065, 1994.
- Kamada, S., et al. J. of Endocrinol. Invest. 16: 425-431, 1993.
- Acosta, T.J. et al. Biol. Reprod. 59: 437-443, 1998.
- Ott (1977) An Introduction to Statistical Methods and Data Analysis. Page 384-388.

ACKNOWLEDGMENTS

- The Lew Wentz Foundation.
- Creekstone Farms for their generous donation of ovaries.
- Lingna Zhang and Marco Albonico for lab assistance.
- The Department of Animal Science at Oklahoma State University.